1	Dose-dependent pro- or anti-fibrotic responses of endometriotic stromal cells to interleukin-1p
2	and tumor necrosis factor α
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4	Running title: Inflammation and endometriosis
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Supplementary Note

Varying doses of IL-1\beta and/or TNFa in the present study

Many previous in vitro experiments used very high levels of IL-1 β and/or TNF α (ng- μ g/mL ranges) to elucidate the roles of these two proinflammatory cytokines in the pathophysiology of endometriosis. However, it is not clear whether such high levels of IL-1 β and/or TNF α could be involved in the pathophysiology of endometriosis. Previous studies reported much lower levels of IL-1 β and/or TNF α in serum and peritoneal fluid of patients with endometriosis (pg/mL range). Although proinflammatory cytokines are produced predominantly by activated macrophages, a previous in vitro study showed that peritoneal macrophages from patients with endometriosis produced TNF α in the pg/ml range following stimulation with LPS. High levels of IL-1 β and/or TNF α (ng- μ g/mL ranges) in previous experiments $^{1-5}$ may not be recapitulated in the in vivo environment of patients with endometriosis.

Thus, first, we measured active IL-1β and TNFα secretion of endometriotic stromal cells, and menstrual endometrial stromal cells of patients with endometriosis and those of healthy control women, after stimulation with LPS or Poly I:C, to estimate potential pathophysiological ranges of IL-1β and TNFα in endometriotic tissues. In endometriosis, as in many other fibrotic disorders, ¹¹ the inflammatory triggers remain unknown. Thus, in the present in vitro experiments, inflammation was non-specifically triggered by stimulation of innate immunity with either lipopolysaccharide (LPS) or polyinosinic:polycytidylic acid (Poly I:C). ¹¹ Both toll-like receptor (TLR)-3 and -4 recognize dying/dead cells. ¹² During menstruation, dying endometrial cells may activate TLRs, including TLR-3 and -4, in endometrial cells. Furthermore, studies have shown that sustained TLR signaling contributes to the development of many chronic inflammatory and autoimmune diseases. ^{13, 14} Thus, in the present study, inflammation was triggered by acute (4-h) and chronic (24-h) stimulation with either LPS or Poly I:C.

43	The present ELISA analyses suggested that TLR activation may induce low-grade local
44	inflammation in endometriotic and menstrual endometrial stromal cells. According to the present
45	ELISA analysis results as well as those of previous in vivo studies, 6-10 we further evaluated the effects
46	of varying ranges of IL-1β and TNFα (pg/mL range) on fibrosis in endometriosis.

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Supplementary Methods

Treatment of cells

For enzyme-linked immunosorbent assay (ELISA) analyses, cells were incubated with either LPS (100 ng/mL), Poly I:C (10 μg/mL), or vehicle alone for 4 h or 24 h. Human recombinant IL-1β protein (R&D Systems, Lille, France) and TNFα protein (R&D Systems) were dissolved in phosphate-buffered saline (PBS) containing 0.1% bovine serum albumin (BSA). LPS (Sigma-Aldrich), Poly I:C (Sigma-Aldrich), or adenosine triphosphate (ATP; Sigma-Aldrich) were dissolved in phenol red-free DMEM:Nutrient Mixture F-12 (DMEM/F-12) (Life Technologies).

Cytokine quantification

Secreted IL-1 β and TNF α levels in culture supernatants were assayed in duplicate and quantified by ELISA with DuoSet ELISA kits (R&D Systems) according to the manufacturer's instructions. The lower detection limits were 1.0 pg/mL (IL-1 β) and 6.23 pg/mL (TNF α). Briefly, cells (3 × 10⁴ cells per well) were plated in 24-well plates. After 48 hours, LPS, Poly I:C, or vehicle were added with 500 μ L culture media (2% charcoal-stripped FBS) (Sigma-Aldrich) and incubated for 4 h or 24 h, and supernatants were collected. We utilized a well-established LPS or Poly I:C (1st signal; priming)/ATP (a damage-associated molecular pattern [DAMP]) (2nd signal; NLR activation) stimulation protocol to measure active IL-1 β secretion after activation of the NLRP3 inflammasome. Cells were incubated with 3 mM ATP for the last 15 min based on the method of Stoffels et al. to measure active IL-1 β secretion. Absorbance was read at 450/540 nm using a Multiskan microplate reader (Thermo Scientific). The concentration (pg/mL) was normalized to total protein content (pg/ μ g total protein) to control for unwanted sources of variation. Intra-assay precision was 4.3% (IL-1 β) and 5.4% (TNF α), and inter-assay precision was 7.4% (IL-1 β) and 8.9% (TNF α).

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Supplementary Results

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121 Effects of LPS or Poly I:C stimulation on IL-1\beta and TNF\alpha secretion in endometrial and 122 endometriotic stromal cells 123 Neither LPS nor Poly I:C stimulation had a significant effect on IL-1β secretion at 4 h in M-EShealthy (Supplementary Figure S1A). In contrast, after 24-h chronic stimulation with either LPS or 124 125 Poly I:C, IL-1β secretion significantly increased in M-ES-healthy (Supplementary Figure S1B). Both 126 LPS and Poly I:C stimulation significantly increased IL-1β secretion in M-ES-endo at 4 h (Supplementary Figure S1A) and 24 h (Supplementary Figure S1B). In both M-ES-healthy and M-ES-127 endo, both LPS and Poly I:C stimulation significantly increased TNFα secretion at 4 h (Supplementary 128 Figure S1C) and 24 h (Supplementary Figure S1D). In ectopic endometriotic cells (Ectopic-ES), 129 130 neither LPS nor Poly I:C stimulation had a significant effect on IL-1β (Supplementary Figure S1A) or TNFα (Supplementary Figure S1C) secretion at 4 h. In contrast, after 24-h chronic stimulation with 131 either LPS or Poly I:C, IL-1β (Supplementary Figure S1B) and TNFα (Supplementary Figure S1D) 132 133 secretion significantly increased.

135	Supplementary Figure Legends
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137	Supplementary Figure S1: Effects of LPS (100 ng/mL) or Poly I:C (10 µg/mL) stimulation for 4
138	h or 24 h on interleukin-1 β (IL-1 β) (A, B) secretion or tumor necrosis factor α (TNFa) (C, D)
139	secretion in menstrual endometrial stromal cells of healthy fertile women (M-ES-healthy) and
140	those of patients with endometriosis (M-ES-endo), and endometriotic stromal cells (Ectopic-ES)
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142	Numerical values are presented as the mean + SD.
143	*: p<0.05 versus control (vehicle alone) within the same group.
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145	M-ES-healthy: n=6
146	M-ES-endo: n=8
147	Ectopic-ES: n=10
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149	Supplementary Figure S2
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151	Effects of IL-1β on cell proliferation of Ectopic-ES derived from deep infiltrating endometriosis
152	versus ovarian endometriosis.
153	Cells were incubated for 48 h at the indicated concentrations.
154	Percent cell proliferation was calculated as percent of vehicle control after 48-h treatment.
155	Numerical values are presented as the mean \pm SD.
156	
157	C: control (vehicle alone)
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159	Deep infiltrating endometriosis: n=8
160	Ovarian endometriosis: n=8

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162	Supplementary Figure S3: Effects of combinations of IL-1 β and TNF α with or without TGF- β 1
163	on mRNA expression of Col I (A, D, G), MMP-1 (B, E, H), and αSMA (C, F, I) in M-ES-healthy
164	(A-C), M-ES-endo (D-F), and Ectopic-ES (G-I).
165	Cells were incubated at the indicated concentrations.
166	Expression levels of Col-I, MMP-1, and αSMA mRNAs are given relative to the expression level of
167	the reference gene, glyceraldehyde 3-phosphate dehydrogenase (GAPDH).
168	Numerical values are presented as the mean + SD.
169	*: p<0.05 versus control (vehicle alone).
170	#: p<0.05 versus TGF-β1 alone
171	
172	Effects of the highest dose of IL-1 β (100 pg/mL) on mRNA expression of Col I, MMP-1, and α SMA
173	in M-ES-healthy and M-ES-endo were excluded for further analyses, due to markedly reduced cell
174	viability.
175	Because there were no significant differences in either Col I or αSMA protein expression between
176	vehicle-treated control after 48h and 96h, results of vehicle-treated control after 96h were not shown.
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178	M-ES-healthy: n=8
179	M-ES-endo: n=16
180	Ectopic-ES: n=22
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182	Supplementary Figure S4: Effects of combinations of IL-1 β and TNF α with or without TGF- β 1
183	on Col I protein expression (A, C, E) and αSMA+ stress fibers (B, D, F) in M-ES-healthy (A, B)
184	M-ES-endo (C, D), and Ectopic-ES (E, F).

(A-F) The percentage of cells with Col I+ or αSMA+ stress fibers after stimulation with IL-1β or 185 TNFα alone or following TGF-β1 5 ng/mL stimulation in M-ES-healthy, M-ES-endo, and Ectopic-ES. 186 187 *: p<0.05: versus control (vehicle alone) 188 #: p<0.05 versus TGF-β1 alone Numerical values are presented as the mean \pm SD. 189 M-ES-healthy: n=8 190 M-ES-endo: n=16 191 192 Ectopic-ES: n=22 Effects of the highest dose of IL-1β (100 pg/mL) on protein expression of Col I and αSMA in M-ES-193 194 healthy and M-ES-endo were excluded for further analyses, due to markedly reduced cell viability. Because there were no significant differences in either Col I or αSMA protein expression between 195 vehicle-treated control after 48h and 96h, results of vehicle-treated control after 96h were not shown. 196 197 198 Supplementary Figure S5: Representative photomicrographs of double immunofluorescence staining 199 for Col I and αSMA in M-ES-healthy, M-ES-endo, and Ectopic-ES after stimulation with IL-1β and/or 200 TNFα alone or following TGF-β1 5 ng/mL stimulation. Scale bar: 50 μm. 201 202 M-ES-healthy: n=8 203 M-ES-endo: n=16 204 Ectopic-ES: n=22 205 206 Effects of the highest dose of IL-1β (100 pg/mL) on protein expression of Col I and αSMA in M-ES-207 healthy and M-ES-endo were excluded for further analyses, due to markedly reduced cell viability.

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